



# *Photobacterium damsela* subsp. *damsela*, a bacterium pathogenic for marine animals and humans

Amable J. Rivas, Manuel L. Lemos and Carlos R. Osorio\*

Institute of Aquaculture, University of Santiago de Compostela, Santiago de Compostela, Spain

## Edited by:

Daniela Ceccarelli, University of Maryland, USA

## Reviewed by:

Devaki Bhaya, Carnegie Institution for Science, USA

Ming-Lun Chen, National Penghu University of Science and Technology, Taiwan

## \*Correspondence:

Carlos R. Osorio, Institute of Aquaculture, Campus Vida, University of Santiago de Compostela, 15782 Santiago de Compostela, Galicia, Spain  
e-mail: cr.osorio@usc.es

*Photobacterium damsela* subsp. *damsela* (formerly *Vibrio damsela*) is a pathogen of a variety of marine animals including fish, crustaceans, molluscs, and cetaceans. In humans, it can cause opportunistic infections that may evolve into necrotizing fasciitis with fatal outcome. Although the genetic basis of virulence in this bacterium is not completely elucidated, recent findings demonstrate that the phospholipase-D Dly (damselysin) and the pore-forming toxins HlyA<sub>pl</sub> and HlyA<sub>ch</sub> play a main role in virulence for homeotherms and poikilotherms. The acquisition of the virulence plasmid pPHDD1 that encodes Dly and HlyA<sub>pl</sub> has likely constituted a main driving force in the evolution of a highly hemolytic lineage within the subspecies. Interestingly, strains that naturally lack pPHDD1 show a strong pathogenic potential for a variety of fish species, indicating the existence of yet uncharacterized virulence factors. Future and deep analysis of the complete genome sequence of *Photobacterium damsela* subsp. *damsela* will surely provide a clearer picture of the virulence factors employed by this bacterium to cause disease in such a varied range of hosts.

**Keywords:** *Photobacterium damsela*, hemolysin, damselysin, hlyA, pore-forming toxin

## PHOTOBACTERIUM DAMSELAE SUBSP. DAMSELAE

*Photobacterium damsela* subsp. *damsela* is a marine bacterium of the family *Vibrionaceae* that causes infections in a variety of marine animals and also in humans. A bit of historic perspective is necessary in order to understand its current taxonomic placement as well as the changes in its nomenclature during the past decades. In 1971 an “unnamed marine *Vibrio*” was isolated as the causative agent of a human infectious case (Morris et al., 1982). Later, this same organism was isolated from skin ulcers of damselfish (*Chromis punctipinnis*) and the name *Vibrio damsela* was first coined (Love et al., 1981). Further genetic and phenotypic studies indicated that the strains of *V. damsela* were closely related to species of the genus *Photobacterium*, and the name *Photobacterium damsela* was proposed (Smith et al., 1991). In 1995, DNA–DNA hybridization data and 16S rRNA sequence analysis demonstrated that *Photobacterium damsela* was closely related to a fish pathogen formerly named *Pasteurella piscicida*, the causative agent of pasteurellosis in fish. Hence, these two organisms were assigned to the same species epithet, *Photobacterium damsela*, with category of subspecies (Gauthier et al., 1995), *Photobacterium damsela* subsp. *damsela* and *Photobacterium damsela* subsp. *piscicida* respectively. Despite their similarity at the 16S gene sequence and the high percentage of DNA–DNA relatedness between them, these two subspecies are clearly distinguished by several phenotypic traits (Fouz et al., 1992; Magariños et al., 1992; Thyssen et al., 1998; Botella et al., 2002). Differential phenotypical tests of interest for subspecies discrimination that are positive only for subsp. *damsela* include motility, nitrate reduction and hemolysis on sheep blood agar. Of special relevance is the ability of most subsp. *damsela* strains to grow at 37°C (a temperature inhibitory for subsp. *piscicida*), a trait that allows *Photobacterium damsela*

subsp. *damsela* to potentially colonize and establish an infection in a homeotherm animal.

## A PATHOGEN OF MARINE ANIMALS

*Photobacterium damsela* subsp. *damsela* is an autochthonous member of aquatic ecosystems. Strains of this pathogen have been isolated from sea and estuarine waters, from seaweeds, from apparently uninfected marine animals (Buck et al., 2006; Serracca et al., 2011) and from seafood (Lozano-León et al., 2003; Chiu et al., 2013), and it is considered a common member of the natural microbiota of healthy carcharhinid sharks (Grimes et al., 1985).

In addition, *Photobacterium damsela* subsp. *damsela* is considered a primary pathogen of several species of wild fish (damselfish, catfish, shark, stingray, etc.), as well as of fish species of economical importance in aquaculture, causing wound infections and hemorrhagic septicemia. Cultivated species reported to be affected by this pathogen include turbot (*Psetta maxima*; Fouz et al., 1992), rainbow trout (*Oncorhynchus mykiss*; Pedersen et al., 2009), ovate pompano (*Trachinotus ovatus*; Zhao et al., 2009), eel (*Anguilla reinhardtii*; Ketterer and Eaves, 1992), sea bream (*Sparus aurata*; Vera et al., 1991), sea bass (*Dicentrarchus labrax*), yellowtail (*Seriola quinqueradiata*), redbanded seabream (*Pagrus auriga*), white seabream (*Diplodus sargus*), and meagre (*Argyrosomus regius*; Labella et al., 2006, 2010a,b), among others. The recent first reports on isolation of this pathogen from diseased marine fish of new cultured species, suggest that *Photobacterium damsela* subsp. *damsela* can be considered as an emerging pathogen in marine aquaculture (Labella et al., 2011).

Moreover, *Photobacterium damsela* subsp. *damsela* has been isolated as a pathogen of brown shark (*Carcharhinus plumbeus*;

Grimes et al., 1984), of reptiles as the leatherback sea turtle (*Dermochelys coriacea*; Obendorf et al., 1987), molluscs (*Octopus joubini*; Hanlon et al., 1984), crustaceans (Song et al., 1993; Vaseeharan et al., 2007), dolphins (*Tursiops truncatus* and *Delphinus delphis*; Fujioka et al., 1988; Buck et al., 1991) and Bryde's whale (*Balaenoptera edeni*; Buck et al., 1991).

Virulent isolates are capable of survival in seawater microcosms at 14–22°C as culturable bacteria for long periods of time, maintaining their infectivity for fish (Fouz et al., 1998). Similarly, this pathogen can infect new fish hosts through water, and the spread of the disease depends largely on water temperature and salinity (Fouz et al., 2000). Typical signs of the disease in infected fish include hemorrhaged areas on the body surface and ulcerative lesions. In damselfish, ulcers typically occur in the region of the pectoral fin and caudal peduncle and may reach 5–20 mm in diameter (Love et al., 1981), while in turbot the most remarkable symptoms are extensive hemorrhages in eyes, mouth, and jaws (Fouz et al., 1995).

Experimental inoculation of *Photobacterium damsela* subsp. *damsela* extracellular products (ECPs) in a redbanded seabream model was reported to cause lethargy, increase in the respiratory frequency, mucus production, presence of ascitic liquid, hemorrhagic and enlarged liver, and hemorrhages in the abdominal cavity (Labella et al., 2010b). A histological analysis of internal organs in experimentally infected turbot indicated that the ECPs and cells of virulent strains cause similar tissue damage (Fouz et al., 1995). Structural changes included destruction and necrosis of cells, as well as accumulation of blood cells in interstitial tissue.

#### ***Photobacterium damsela* subsp. *damsela* AS A HUMAN PATHOGEN**

Most of the reported infections caused by *Photobacterium damsela* subsp. *damsela* in humans have their primary origin in wounds exposed to salt or brackish water, inflicted during fish and tools handling (Morris et al., 1982; Dryden et al., 1989; Yuen et al., 1993; Shin et al., 1996; Tang and Wong, 1999; Barber and Swygert, 2000; Goodell et al., 2004; Aigbivhalu and Maraqa, 2009). Unusual cases of infection after ingestion of raw seafood (Kim et al., 2009) and through the urinary tract by exposure to sea water (Alvarez et al., 2006) were also reported. The majority of the cases occurred in coastal areas of the United States of America, Australia, and Japan.

*Photobacterium damsela* subsp. *damsela* can cause an extreme variant of a highly severe necrotizing fasciitis, and antibiotic administration proved unable to control the progression of fatal infections in some cases (Clarridge and Zigelboimdaum, 1985; Fraser et al., 1997; Yamane et al., 2004). It is interesting to note that some authors recommend to surgically debride and amputate without hesitation at a very early point of the infection by *Photobacterium damsela* subsp. *damsela*, to save the lives of patients (Goodell et al., 2004). Some patients infected by *Photobacterium damsela* subsp. *damsela* developed multiple organ failure within a few hours from the onset of initial symptoms, despite intensive chemotherapy and surgical treatments. As an example, in a fatal case reported in 1984 in which a patient injured his hand while handling a catfish, bulle formation occurred on the hand and a marked edema extended through the forearm in less than 24 h (Clarridge and Zigelboimdaum, 1985), and although the

affected area was extensively debrided the patient died after a series of complications. The bacterium was recovered in high numbers from the tissue sample but only in very small numbers from the bulle fluid. In another fatal case reported in a patient injured while handling fish, *Photobacterium damsela* subsp. *damsela* was isolated in pure culture from wound specimens but failed to be isolated from blood samples (Fraser et al., 1997). These observations prompted these authors to suggest that a virulence factor or systemic toxin released by this bacterium contributed to the tissue damage and to the fatal outcome, rather than the septicemia itself. However, in other clinical cases this pathogen was recovered from blood (Perez-Tirse et al., 1993; Shin et al., 1996; Yamane et al., 2004).

Necrotizing fasciitis due to *Photobacterium damsela* subsp. *damsela* demonstrates more serious complications and a higher mortality rate than that caused by *Vibrio vulnificus*. While *V. vulnificus* usually affects persons with underlying diseases (as chronic liver disease and diabetes mellitus), necrotizing fasciitis by *Photobacterium damsela* subsp. *damsela* sometimes occurs in healthy hosts (Morris et al., 1982; Perez-Tirse et al., 1993; Yuen et al., 1993).

#### **VIRULENCE FACTORS**

##### ***Iron uptake systems***

Early studies reported that *Photobacterium damsela* subsp. *damsela* can utilize heme, hemoglobin and ferric ammonium citrate as sole iron sources *in vitro* (Fouz et al., 1994). The complete sequence of 10 genes encoding a system for the utilization of heme as iron source was described in a human isolate of *Photobacterium damsela* subsp. *damsela*, and cloning of the complete system into *E. coli* conferred to this species the ability to use hemin and hemoglobin as iron sources (Rio et al., 2005). The presence of the heme receptor gene *hutA* was demonstrated in subsp. *damsela* isolates from fish and humans, and the identity at the DNA sequence level between the heme uptake clusters of subsp. *damsela* and subsp. *piscicida* strains was 97% (Rio et al., 2005). Although no functional studies were conducted with the heme uptake genes of subsp. *damsela*, it was recently demonstrated that this cluster is essential for heme utilization in subsp. *piscicida*, and two genes of a hemin ABC-transporter proved to be expressed during the infective process in a fish model (Osorio et al., 2010). Actually, an increase in the susceptibility of both fish and mice to infection by virulent *Photobacterium damsela* subsp. *damsela* strains in virulence assays conducted with iron-overloaded animals had been demonstrated in former studies (Fouz et al., 1994). It is also known that this bacterium produces a hydroxamate-type siderophore, and the synthesis of several high-molecular weight outer membrane proteins induced under iron limitation conditions was reported (Fouz et al., 1997). Although the precise chemical structure of the siderophore(s) is so far unknown, recent unpublished work from our laboratory demonstrated that vibrioferrin is being produced by some strains.

##### ***Cytotoxins with hemolytic activity***

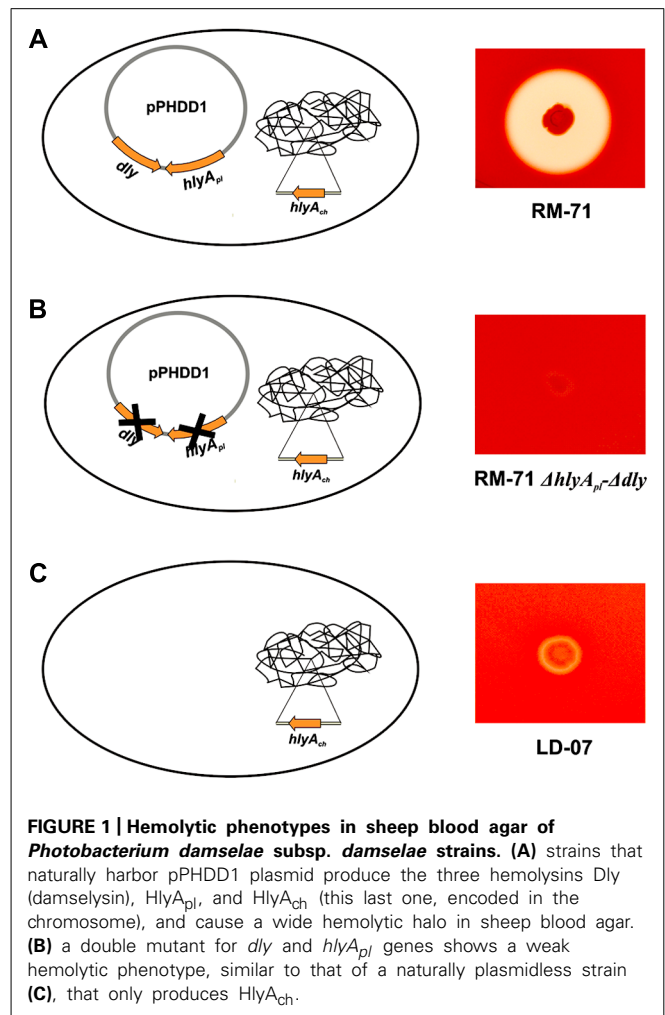
Pioneering studies (Kreger, 1984) reported the existence of a correlation between the ability of *Photobacterium damsela* subsp.

*damsela* isolates to cause disease in mice and the production of large amounts of a heat-labile cytolytic toxin *in vitro*. Later, the same authors purified a toxin that exhibited strong hemolytic activity against erythrocytes of a variety of animal species (Kothary and Kreger, 1985). In subsequent studies, this toxin named damselysin (Dly) was defined as a phospholipase-D active against sphingomyelin, with hemolytic activity (Kreger et al., 1987), and its gene (*dly*) was cloned and sequenced (Cutter and Kreger, 1990). Dly was thus considered to be the main virulence factor of *Photobacterium damsela* subsp. *damsela* for mice. Further studies reported the existence of hemolytic strains of *Photobacterium damsela* subsp. *damsela* that tested negative for *dly* gene, which suggested that Dly was not the only hemolysin in the subspecies (Osorio et al., 2000). It was also demonstrated by several authors that presence of *dly* is not a prerequisite for the hemolytic activity and for the pathogenicity for mice or fish, since *dly* negative strains bear virulence potential for animals and also toxicity for homeotherm and poikilotherm cell lines (Osorio et al., 2000; Labella et al., 2010b).

The genomic context of *dly* gene remained uncharacterized for decades, and it was initially proposed that it could be carried on a mobile or unstable DNA element (Cutter and Kreger, 1990). Recently, the authors' laboratory identified and characterized a 150 kb plasmid, which was dubbed pPHDD1, that contains the genes for Dly as well as for a HlyA toxin of the pore-forming toxin family (Rivas et al., 2011). Only a fraction of *Photobacterium damsela* subsp. *damsela* strains harbor pPHDD1, and these strains exhibit a much wider hemolytic halo on sheep blood agar plates than the plasmidless strains (Figure 1). Interestingly, pPHDD1 occurs in both fish and human isolates and it is not restricted to a unique animal host species (Rivas et al., 2011). In addition to being necessary to cause strong hemolytic haloes on blood agar plates, the two pPHDD1-encoded hemolysins play a crucial role in virulence for fish and mice in strains that naturally harbor the plasmid. Hence, mutation of both *dly* and *hlyA* genes in a pPHDD1-harboring strain renders the strain non-virulent for fish, and only slightly virulent for mice (Table 1), and the hemolytic phenotype on sheep blood agar of a *dly* and *hlyA* double mutant resembles that of naturally plasmidless strains (Figure 1; Rivas et al., 2011, 2013).

The hemolytic activity exhibited by plasmidless strains was recently demonstrated to be caused by a chromosome-encoded *hlyA* gene, which was dubbed *hlyA<sub>ch</sub>* in order to differentiate it from the plasmid *hlyA* gene (hereafter *hlyA<sub>pl</sub>*; Rivas et al., 2013). It was found that all the hemolytic *Photobacterium damsela* subsp. *damsela* strains harbor *hlyA<sub>ch</sub>* gene, which is the only hemolytic determinant in plasmidless strains. Thus, pPHDD1-harboring isolates produce three different hemolysins. In hemolytic assays carried out with bacterial ECPs and with sheep erythrocytes, it was demonstrated that Dly acts in a synergistic manner with HlyA<sub>pl</sub> and HlyA<sub>ch</sub>, whereas the effect between HlyA<sub>pl</sub> and HlyA<sub>ch</sub> showed to be additive but not synergistic (Rivas et al., 2013).

Although each of the three hemolysins individually proved to be sufficient to cause death in mice, each one contributes to virulence in a different degree. The contribution of HlyA<sub>ch</sub> to virulence for mice is the lowest among the three toxins. Altogether, albeit



**FIGURE 1 | Hemolytic phenotypes in sheep blood agar of *Photobacterium damsela* subsp. *damsela* strains. (A)** strains that naturally harbor pPHDD1 plasmid produce the three hemolysins Dly (damselysin), HlyA<sub>pl</sub>, and HlyA<sub>ch</sub> (this last one, encoded in the chromosome), and cause a wide hemolytic halo in sheep blood agar. **(B)** a double mutant for *dly* and *hlyA<sub>pl</sub>* genes shows a weak hemolytic phenotype, similar to that of a naturally plasmidless strain **(C)**, that only produces HlyA<sub>ch</sub>.

**Table 1 | Role of the three *Photobacterium damsela* subsp. *damsela* hemolysins in virulence for mice and fish (turbot).**

| Strain   | Hemolysin(s) produced                     | Number of dead mice (n = 15) | Number of dead fish (n = 15) |
|--|---|------------------------------|------------------------------|
| Parental   | HlyA <sub>ch</sub> HlyA <sub>pl</sub> Dly | 15                           | 15                           |
| Δ <i>hlyA<sub>ch</sub></i>   | HlyA <sub>pl</sub> Dly                    | 13                           | 9*                           |
| Δ <i>hlyA<sub>pl</sub></i>   | HlyA <sub>ch</sub> Dly                    | 10*                          | 14                           |
| Δ <i>dly</i>   | HlyA <sub>ch</sub> HlyA <sub>pl</sub>     | 10*                          | 4*                           |
| Δ <i>hlyA<sub>ch</sub></i> Δ <i>hlyA<sub>pl</sub></i>              | Dly                                       | 12                           | 5*                           |
| Δ <i>hlyA<sub>ch</sub></i> Δ <i>dly</i>                            | HlyA <sub>pl</sub>                        | 9*                           | 0*                           |
| Δ <i>hlyA<sub>pl</sub></i> Δ <i>dly</i>                            | HlyA <sub>ch</sub>                        | 3*                           | 0*                           |
| Δ <i>hlyA<sub>ch</sub></i> Δ <i>hlyA<sub>pl</sub></i> Δ <i>dly</i> | none                                      | 0*                           | 0*                           |

Parental and mutant strains were inoculated in groups of 15 animals, at doses of  $2.1 \times 10^6$  bacterial cells per mouse and  $2.1 \times 10^4$  bacterial cells per fish. The number of dead animals out of the total number of inoculated animals (15) is indicated. Asterisks denote that significant differences exist between a given mutant and the parental strain, using U-test (\* $P < 0.05$ ).

the highest values of mortality for mice are achieved only when the three hemolysins are being produced, Dly and HlyA<sub>pl</sub> demonstrated to be main contributors in the virulence of *Photobacterium damsela* subsp. *damsela* for mice (Rivas et al., 2013; **Table 1**).

Interestingly, the contribution of each hemolysin to virulence was found to vary depending on whether the host animal tested was mouse or turbot. When virulence experiments were conducted with turbot, it was found that among all the hemolysin gene mutants only the Dly-producing strains caused death in fish. This finding demonstrated that any of the two HlyA alone does not cause death in turbot, but rather one of the two HlyA needs the presence of either Dly or the other HlyA to cause death in fish. The production of Dly in combination with any of the two HlyA caused an increase in the number of dead fish with respect to the production of Dly alone, and this increase was found to be particularly evident when Dly was combined with HlyA<sub>ch</sub>. This clearly suggests that, unlike what is observed in mice, the contribution of hemolysins to virulence for fish is not so much based on the individual effects of each hemolysin but rather on the combined (synergistic) effects between Dly and HlyA (Rivas et al., 2013; **Table 1**). These findings also state the importance of pPHDD1 plasmid in virulence for fish, since Dly is necessary for the synergistic effect.

#### **Other exoenzymes and exotoxins: toxicity of the extracellular products**

Early studies detected several enzymatic activities in the ECPs of *Photobacterium damsela* subsp. *damsela*, which included phospholipase and hemolysin activities (Fouz et al., 1993). More recent data confirmed that the ECPs of *Photobacterium damsela* subsp. *damsela* are strongly lethal for fish, and enzymatic activities such as amylase, lipase, phospholipase, alkaline phosphatase, esterase–lipase, acid phosphatase, and β-glucosaminidase were evidenced (Labella et al., 2010b). Moreover, treatment at 100°C for 10 min of the ECPs abolished the ability to cause death in fish, suggesting that toxicity was not due to the thermoresistant lipopolysaccharide content. *Photobacterium damsela* subsp. *damsela* ECPs also displayed cytotoxic activity for different fish and mammalian cell lines (Wang et al., 1998; Labella et al., 2010b). Different studies found a correlation between virulence of the strain and toxicity of the ECPs, with toxicity being limited to ECPs from strains that were also virulent for fish (Fouz et al., 1995; Labella et al., 2010b). Of maximum interest is the observation that strains lacking pPHDD1 plasmid and thus being negative for *dly* and *hlyA<sub>pl</sub>* genes, are virulent for fish and their ECPs are cytotoxic for cell lines. In addition, a comprehensive study reported that none of the enzymatic activities detected in the *Photobacterium damsela* subsp. *damsela* ECPs could be related with the degree of toxicity either *in vivo* or *in vitro* (Labella et al., 2010b). Most *Photobacterium damsela* subsp. *damsela*

strains test negative for protease activities as caseinase and gelatinase (Fouz et al., 1992; Labella et al., 2010b). This suggests that other, yet uncharacterized molecules produced by *Photobacterium damsela* subsp. *damsela* cells play a role in toxicity for animals and for cell lines. In this regard, previous studies detected the existence of an acetylcholinesterase activity (ictiotoxin) with neurotoxic activity in several species of *Vibrionaceae*, including *Photobacterium damsela* subsp. *damsela* (Perez et al., 1998), although the genetic basis for this neurotoxic activity remains unknown.

#### **FUTURE PERSPECTIVES**

An interesting observation that remains to be explained at the genetic level is the finding that plasmidless *Photobacterium damsela* subsp. *damsela* strains are virulent for fish and toxic for homeotherm and poikilotherm cell lines (Fouz et al., 1993; Osorio et al., 2000; Labella et al., 2010b, 2011). Since plasmidless strains lack *dly* and *hlyA<sub>pl</sub>* genes, and since *dly hlyA<sub>pl</sub>* double mutants are significantly reduced in its virulence for both mice and fish (Rivas et al., 2013), it is evident that plasmidless strains encode virulence factors that either are not encoded by pPHDD1-harboring strains or their expression is repressed in presence of pPHDD1-encoded genes.

The recent completion of the genome sequence of the type strain (ATCC 33539) of this subspecies (deposited in GenBank database in several separate contigs, under accession number ADBS00000000), allows an *in silico* analysis to search for candidate genes encoding potential toxins and other virulence factors. The type strain harbors pPHDD1 plasmid, and preliminary analyses also indicated the presence of genes encoding a type III hemolysin (open reading frame number: VDA003208), and a putative murine toxin (VDA000322) among others. The existence of yet uncharacterized plasmids is also evidenced in the complete genome of ATCC 33539. Studies to functionally characterize novel plasmid content and candidate virulence genes of *Photobacterium damsela* subsp. *damsela* strains are currently under way. It is expected that a deep analysis of the complete genome sequence of *Photobacterium damsela* subsp. *damsela* strains with different isolation origins and virulence properties will provide a clearer picture of the virulence factors employed by this bacterium to cause disease in such a varied range of hosts.

#### **ACKNOWLEDGMENTS**

The work in the authors' laboratory is supported by grant EM2012/043 from Xunta de Galicia, Spain; and by grants AGL2012-39274-C02-01, from the Ministry of Economy and Competitiveness (MINECO) of Spain and CSD2007-00002 (Consolider Aquagenomics) from the Ministry of Science and Innovation (MICINN) of Spain, both (cofunded by the FEDER Programme from the European Union).

#### **REFERENCES**

- Aigbivhalu, L., and Maraqa, N. (2009). *Photobacterium damsela* wound infection in a 14-year-old surfer. *South. Med. J.* 102, 425–426. doi: 10.1097/SMJ.0b013e31819b9491
- Alvarez, J. R., Lamba, S., Dyer, K. Y., and Apuzzio, J. J. (2006). An unusual case of urinary tract infection in a pregnant woman with *Photobacterium damsela*. *Infect. Dis. Obstet. Gynecol.* 2006, 80682. doi: 10.1155/IDOG/2006/80682
- Barber, G. R., and Swygert, J. S. (2000). Necrotizing fasciitis due to *Photobacterium damsela* in a man lashed by a stingray. *N. Engl. J. Med.* 342, 824. doi: 10.1056/NEJM200003163421118
- Botella, S., Pujalte, M. J., Macian, M. C., Ferrus, M. A., Hernandez, J., and Garay, E. (2002). Amplified fragment length polymorphism (AFLP) and biochemical typing of *Photobacterium damsela* subsp. *damsela*. *J. Appl. Microbiol.* 93,

- 681–688. doi: 10.1046/j.1365-2672.2002.01748.x
- Buck, J. D., Overstrom, N. A., Patton, G. W., Anderson, H. F., and Gorzelany, J. F. (1991). Bacteria associated with stranded cetaceans from the Northeast USA and Southwest Florida Gulf coasts. *Dis. Aquat. Org.* 10, 147–152. doi: 10.3354/dao010147
- Buck, J. D., Wells, R. S., Rhinehart, H. L., and Hansen, L. J. (2006). Aerobic microorganisms associated with free-ranging bottlenose dolphins in coastal Gulf of Mexico and Atlantic Ocean waters. *J. Wildl. Dis.* 42, 536–544.
- Chiu, T.-H., Kao, L.-Y., and Chen, M.-L. (2013). Antibiotic resistance and molecular typing of *Photobacterium damsela* subsp. *damsela*, isolated from seafood. *J. Appl. Microbiol.* 114, 1184–1192. doi: 10.1111/jam.12104
- Clarridge, J. E., and Zigelboimdaum, S. (1985). Isolation and characterization of 2 hemolytic phenotypes of *Vibrio damsela* associated with a fatal wound infection. *J. Clin. Microbiol.* 21, 302–306.
- Cutter, D. L., and Kreger, A. S. (1990). Cloning and expression of the damselysin gene from *Vibrio damsela*. *Infect. Immun.* 58, 266–268.
- Dryden, M., Legarde, M., Gottlieb, T., Brady, L., and Ghosh, H. K. (1989). *Vibrio damsela* wound infections in Australia. *Med. J. Aust.* 151, 540–541.
- Fouz, B., Barja, J. L., Amaro, C., Rivas, C., and Toranzo, A. E. (1993). Toxicity of the extracellular products of *Vibrio damsela* isolated from diseased fish. *Curr. Microbiol.* 27, 341–347. doi: 10.1007/BF01568958
- Fouz, B., Biosca, E. G., and Amaro, C. (1997). High affinity iron-uptake systems in *Vibrio damsela*: role in the acquisition of iron from transferrin. *J. Appl. Microbiol.* 82, 157–167.
- Fouz, B., Larsen, J. L., Nielsen, B., Barja, J. L., and Toranzo, A. E. (1992). Characterization of *Vibrio damsela* strains isolated from turbot *Scophthalmus maximus* in Spain. *Dis. Aquat. Org.* 12, 155–166. doi: 10.3354/dao012155
- Fouz, B., Novoa, B., Toranzo, A. E., and Figueras, A. (1995). Histopathological lesions caused by *Vibrio damsela* in cultured turbot, *Scophthalmus maximus* (L) – inoculations with live cells and extracellular products. *J. Fish Dis.* 18, 357–364. doi: 10.1111/j.1365-2761.1995.tb00312.x
- Fouz, B., Toranzo, A. E., Biosca, E. G., Mazoy, R., and Amaro, C. (1994). Role of iron in the pathogenicity of *Vibrio damsela* for fish and mammals. *FEMS Microbiol. Lett.* 121, 181–188. doi: 10.1111/j.1574-6968.1994.tb07097.x
- Fouz, B., Toranzo, A. E., Marco-Noales, E., and Amaro, C. (1998). Survival of fish-virulent strains of *Photobacterium damsela* subsp. *damsela* in seawater under starvation conditions. *FEMS Microbiol. Lett.* 168, 181–186. doi: 10.1111/j.1574-6968.1998.tb13271.x
- Fouz, B., Toranzo, A. E., Milan, M., and Amaro, C. (2000). Evidence that water transmits the disease caused by the fish pathogen *Photobacterium damsela* subsp. *damsela*. *J. Appl. Microbiol.* 88, 531–535. doi: 10.1046/j.1365-2672.2000.00992.x
- Fraser, S. L., Purcell, B. K., Delgado, B., Baker, A. E., and Whelen, A. C. (1997). Rapidly fatal infection due to *Photobacterium (Vibrio) damsela*. *Clin. Infect. Dis.* 25, 935–936. doi: 10.1086/597647
- Fujioka, R. S., Greco, S. B., Cates, M. B., and Schroeder, J. P. (1988). *Vibrio damsela* from wounds in bottlenose dolphins *Tursiops truncatus*. *Dis. Aquat. Org.* 4, 1–8. doi: 10.3354/dao004001
- Gauthier, G., Lafay, B., Ruimy, R., Breittmayer, V., Nicolas, J. L., Gauthier, M., et al. (1995). Small-subunit ribosomal-RNA sequences and whole DNA relatedness concur for the reassignment of *Pasteurella piscicida* (Sniezko et al.) Janssen and Surgalla to the genus *Photobacterium* as *Photobacterium damsela* subsp. *piscicida* comb. nov. *Int. J. Syst. Bacteriol.* 45, 139–144. doi: 10.1099/00207713-45-1-139
- Goodell, K. H., Jordan, M. R., Graham, R., Cassidy, C., and Nasraway, S. A. (2004). Rapidly advancing necrotizing fasciitis caused by *Photobacterium (Vibrio) damsela*: a hyperaggressive variant. *Crit. Care Med.* 32, 278–281. doi: 10.1097/01.CCM.0000104920.01254.82
- Grimes, D. J., Brayton, P., Colwell, R. R., and Gruber, H. (1985). *Vibrios* as autochthonous flora of neritic sharks. *Syst. Appl. Microbiol.* 6, 221–226. doi: 10.1016/S0723-2020(85)80056-4
- Grimes, D. J., Colwell, R. R., Stemmler, J., Hada, H., Maneval, D., Hetrick, F. M., et al. (1984). *Vibrio* species as agents of elasmobranch disease. *Helgol. Meeresunters* 37, 309–315. doi: 10.1007/BF01989313
- Hanlon, R. T., Forsythe, J. W., Cooper, K. M., Dinuzzo, A. R., Folse, D. S., and Kelly, M. T. (1984). Fatal penetrating skin ulcers in laboratory-reared octopuses. *J. Invertebr. Pathol.* 44, 67–83. doi: 10.1016/0022-2011(84)90047-8
- Ketterer, P. J., and Eaves, L. E. (1992). Deaths in captive eels (*Anguilla reinhardtii*) due to *Photobacterium (Vibrio) damsela*. *Aust. Vet. J.* 69, 203–204. doi: 10.1111/j.1751-0813.1992.tb07528.x
- Kim, H. R., Kim, J. W., Lee, M. K., and Kim, J. G. (2009). Septicemia progressing to fatal hepatic dysfunction in a cirrhotic patient after oral ingestion of *Photobacterium damsela*: a case report. *Infection* 37, 555–556. doi: 10.1007/s15010-009-9049-8
- Kothary, M. H., and Kreger, A. S. (1985). Purification and characterization of an extracellular cytolytic produced by *Vibrio damsela*. *Infect. Immun.* 49, 25–31.
- Kreger, A. S. (1984). Cytolytic activity and virulence of *Vibrio damsela*. *Infect. Immun.* 44, 326–331.
- Kreger, A. S., Bernheimer, A. W., Etkin, L. A., and Daniel, L. W. (1987). Phospholipase D activity of *Vibrio damsela* cytolytic and its interaction with sheep erythrocytes. *Infect. Immun.* 55, 3209–3212.
- Labella, A., Berbel, C., Manchado, M., Castro, D., and Borrego, J. J. (2011). “*Photobacterium damsela* subsp. *damsela*, an emerging pathogen affecting new cultured marine fish species in southern Spain,” in *Recent Advances in Fish Farms*, eds Faruk Aral and Zafer Dogu (New York: InTech), 135–152.
- Labella, A., Manchado, M., Alonso, M. C., Castro, D., Romalde, J. L., and Borrego, J. J. (2010a). Molecular intraspecific characterization of *Photobacterium damsela* ssp. *damsela* strains affecting cultured marine fish. *J. Appl. Microbiol.* 108, 2122–2132.
- Labella, A., Sanchez-Montes, N., Berbel, C., Aparicio, M., Castro, D., Manchado, M., et al. (2010b). Toxicity of *Photobacterium damsela* subsp. *damsela* strains isolated from new cultured marine fish. *Dis. Aquat. Org.* 92, 31–40. doi: 10.3354/dao02275
- Labella, A., Vida, M., Alonso, M. C., Infante, C., Cardenas, S., López-Romalde, S., et al. (2006). First isolation of *Photobacterium damsela* ssp. *damsela* from cultured redbanded seabream, *Pagrus auriga* Valenciennes, in Spain. *J. Fish Dis.* 29, 175–179. doi: 10.1111/j.1365-2761.2006.00697.x
- Love, M., Teebkenfisher, D., Hose, J. E., Farmer, J. J., Hickman, F. W., and Fanning, G. R. (1981). *Vibrio damsela*, a marine bacterium, causes skin ulcers on the damselfish *Chromis punctipinnis*. *Science* 214, 1139–1140. doi: 10.1126/science.214.4525.1139
- Lozano-León, A., Osorio, C. R., Núñez, S., Martínez-Urtaza, J., and Magariños, B. (2003). Occurrence of *Photobacterium damsela* subsp. *damsela* in bivalve molluscs from Northwest Spain. *Bull. Eur. Assoc. Fish Pathol.* 23, 40–44.
- Magariños, B., Romalde, J. L., Bandín, I., Fouz, B., and Toranzo, A. E. (1992). Phenotypic, antigenic, and molecular characterization of *Pasteurella piscicida* strains isolated from fish. *Appl. Environ. Microbiol.* 58, 3316–3322.
- Morris, J. G., Wilson, R., Hollis, D. G., Weaver, R. E., Miller, H. G., Tacket, C. O., et al. (1982). Illness caused by *Vibrio damsela* and *Vibrio cholerae*. *Lancet* 319, 1294–1297. doi: 10.1016/S0140-6736(82)92853-7
- Obendorf, D. L., Carson, J., and Mcmanus, T. J. (1987). *Vibrio damsela* infection in a stranded leatherback turtle (*Dermodochelys coriacea*). *J. Wildl. Dis.* 23, 666–668.
- Osorio, C. R., Juiz-Río, S., and Lemos, M. L. (2010). The ABC transporter *hutCD* genes of *Photobacterium damsela* subsp. *piscicida* are essential for haem utilization as iron source and are expressed during infection in fish. *J. Fish Dis.* 33, 649–655. doi: 10.1111/j.1365-2761.2010.01169.x
- Osorio, C. R., Romalde, J. L., Barja, J. L., and Toranzo, A. E. (2000). Presence of phospholipase-D (ply) gene coding for damselysin production is not a pre-requisite for pathogenicity in *Photobacterium damsela* subsp. *damsela*. *Microb. Pathog.* 28, 119–126. doi: 10.1006/mpat.1999.0330
- Pedersen, K., Skall, H. F., Lassen-Nielsen, A. M., Bjerrum, L., and Olesen, N. J. (2009). *Photobacterium damsela* subsp. *damsela*, an emerging pathogen in Danish rainbow trout, *Oncorhynchus mykiss* (Walbaum), mariculture. *J. Fish Dis.* 32, 465–472. doi: 10.1111/j.1365-2761.2009.01041.x
- Perez, M. J., Rodriguez, L. A., and Nieto, T. P. (1998). The acetylcholinesterase ichthyotoxin is a common component in the extracellular products of *Vibrionaceae* strains. *J. Appl. Microbiol.* 84, 47–52. doi: 10.1046/j.1365-2672.1997.00311.x
- Perez-Tirse, J., Levine, J. F., and Mecca, M. (1993). *Vibrio damsela* – a cause of fulminant septicemia. *Arch. Intern. Med.* 153, 1838–1840. doi: 10.1001/archinte.1993.00410150128012
- Río, S. J., Osorio, C. R., and Lemos, M. L. (2005). Heme uptake genes in human and fish isolates of *Photobacterium damsela*: existence of *hutA* pseudogenes. *Arch. Microbiol.* 183, 347–358. doi: 10.1007/s00203-005-0779-4
- Rivas, A. J., Balado, M., Lemos, M. L., and Osorio, C. R. (2011). The *Photobacterium damsela* subsp. *damsela*

- hemolysins damselysin and HlyA are encoded within a new virulence plasmid. *Infect. Immun.* 79, 4617–4627. doi: 10.1128/IAI.05436-11
- Rivas, A. J., Balado, M., Lemos, M. L., and Osorio, C. R. (2013). Synergistic and additive effects of chromosomal and plasmid-encoded hemolysins contribute to hemolysis and virulence in *Photobacterium damsela* subsp. *damsela*. *Infect. Immun.* 81, 3287–3299. doi: 10.1128/IAI.00155-13
- Serracca, L., Ercolini, C., Rossini, L., Battistini, R., Giorgi, I., and Prearo, M. (2011). Occurrence of both subspecies of *Photobacterium damsela* in mullets collected in the river Magra (Italy). *Can. J. Microbiol.* 57, 437–440. doi: 10.1139/w11-021
- Shin, J. H., Shin, M. G., Suh, S. P., Ryang, D. W., Rew, J. S., and Nolte, F. S. (1996). Primary *Vibrio damsela* septicemia. *Clin. Infect. Dis.* 22, 856–857. doi: 10.1093/clinids/22.5.856
- Smith, S. K., Sutton, D. C., Fuerst, J. A., and Reichelt, J. L. (1991). Evaluation of the genus *Listonella* and reassignment of *Listonella damsela* (Love et al.) MacDonell and Colwell to the genus *Photobacterium* as *Photobacterium damsela* comb. nov. with an emended description. *Int. J. Syst. Bacteriol.* 41, 529–534. doi: 10.1099/00207713-41-4-529
- Song, Y. L., Cheng, W., and Wang, C. H. (1993). Isolation and characterization of *Vibrio damsela* infectious for cultured shrimp in Taiwan. *J. Invertebr. Pathol.* 61, 24–31. doi: 10.1006/jipa.1993.1005
- Tang, W. M., and Wong, J. W. K. (1999). Necrotizing fasciitis caused by *Vibrio damsela*. *Orthopedics* 22, 443–444.
- Thyssen, A., Grisez, L., van Houdt, R., and Ollevier, F. (1998). Phenotypic characterization of the marine pathogen *Photobacterium damsela* subsp. *piscicida*. *Int. J. Syst. Bacteriol.* 48, 1145–1151. doi: 10.1099/00207713-48-4-1145
- Vaseeharan, B., Sundararaj, S., Murugan, T., and Chen, J. C. (2007). *Photobacterium damsela* ssp. *damsela* associated with diseased black tiger shrimp *Penaeus monodon* Fabricius in India. *Lett. Appl. Microbiol.* 45, 82–86. doi: 10.1111/j.1472-765X.2007.02139.x
- Vera, P., Navas, J. I., and Fouz, B. (1991). First isolation of *Vibrio damsela* from seabream (*Sparus aurata*). *Bull. Eur. Assoc. Fish Pathol.* 11, 112–113.
- Wang, X. H., Oon, H. L., Ho, G. W. P., Wong, W. S. F., Lim, T. M., and Leung, K. Y. (1998). Internalization and cytotoxicity are important virulence mechanisms in vibrio-fish epithelial cell interactions. *Microbiology* 144, 2987–3002. doi: 10.1099/00221287-144-11-2987
- Yamane, K., Asato, J., Kawade, N., Takahashi, H., Kimura, B., and Arakawa, Y. (2004). Two cases of fatal necrotizing fasciitis caused by *Photobacterium damsela* in Japan. *J. Clin. Microbiol.* 42, 1370–1372. doi: 10.1128/JCM.42.3.1370-1372.2004
- Yuen, K. Y., Ma, L., Wong, S. S. Y., and Ng, W. F. (1993). Fatal necrotizing fasciitis due to *Vibrio damsela*. *Scand. J. Infect. Dis.* 25, 659–661. doi: 10.3109/00365549309008557
- Zhao, D. H., Sun, J. J., Liu, L., Zhao, H. H., Wang, H. F., Liang, L. Q., et al. (2009). Characterization of two phenotypes of *Photobacterium damsela* subsp. *damsela* isolated from diseased juvenile *Trachinotus ovatus* reared in cage mariculture. *J. World Aquac. Soc.* 40, 281–289. doi: 10.1111/j.1749-7345.2009.00251.x

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 23 July 2013; paper pending published: 12 August 2013; accepted: 02 September 2013; published online: 25 September 2013.

Citation: Rivas AJ, Lemos ML and Osorio CR (2013) *Photobacterium damsela* subsp. *damsela*, a bacterium pathogenic for marine animals and humans. *Front. Microbiol.* 4:283. doi: 10.3389/fmicb.2013.00283

This article was submitted to *Aquatic Microbiology*, a section of the journal *Frontiers in Microbiology*.

Copyright © 2013 Rivas, Lemos and Osorio. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.